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Release and Recovery of Introduced Parasites of the Alfalfa Weevil in Eastern North America

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Washington, D.C. Issued August 1976

Release and Recovery of Introduced Parasites of the Alfalfa Weevil in Eastern North America

By Richard J. Dysart and William H. Day, research entomologists, Beneficial Insects Research Laboratory, Northeastern Region, Agricultural Research Service

The alfalfa weevil (*Hypera postica* (Gyllenhal)), an introduced pest of European or Eurasian origin, was first detected in Utah in 1904. In 1951, a new invasion of this insect was discovered in Maryland, and this eastern population, hereinafter called "the eastern strain," has gradually moved into

all alfalfa-growing areas of Eastern North America. As a result of both infestations, the alfalfa weevil is now in all the 48 contiguous States and in 4 Canadian Provinces. It is considered to be the most important single pest of alfalfa in North America.

PURPOSE AND SCOPE

In 1957, personnel of the Beneficial Insects Research Laboratory at Moorestown, N.J. (now at Newark, Del.) began inoculative releases of imported parasites against the eastern strain of the alfalfa weevil. The objective of this biological control work is to distribute and establish the exotic natural enemies throughout the range of the eastern strain and thus reduce the weevil population to the approximate level that occurs in Europe. In cooperation with research workers in many States, thousands of releases and recoveries have been made of 13 parasite species in this massive and continuing program.

This publication has three sections. In the first is a brief account of each parasite or predator of the alfalfa weevil that has been established, liberated, or is known to occur in Eastern North America. These descriptions are arranged according to the host stage attacked. In the second are suggested methods for parasite releases and recovery attempts. In the third are listed all releases and recoveries ¹ of introduced parasites on the

eastern strain of the alfalfa weevil from 1957 through 1975. In that respect this report is a revision of and a supplement to a previous publication.²

This report provides a detailed record of liberations and recoveries so that biocontrol workers will be able to relate colonization efforts in their particular States with those in the surrounding areas. We hope that the release data summarized here will encourage entomologists to search for parasite species that may be successfully established in their areas but that have not yet been recovered.

The geographical region of North America covered in this publication is east of longitude 100° W. The 100th meridian was selected because it approximates the point where the eastern and western strains of the weevil converged in 1971. Although the colonization of parasites on the western strain of the alfalfa weevil is beyond the scope of this report, we have listed in the appendix all western releases since 1957 where the parasites were supplied by the U.S. Department of Agriculture.

¹ In this report, "recovery" of a parasite species means that it has been collected in the field or reared or dissected from host material obtained from the release site or elsewhere at least 1 year after the release. Survival in the field for this length of time usually indicates that the species has become permanently established in a given area.

² Brunson, M. H., and Coles, L. W. The introduction, release, and recovery of parasites of the alfalfa weevil in eastern united states. U.S. Dept. Agr. Prod. Res. Rpt. 101, 12 pp. 1968.

DESCRIPTIONS OF PARASITES

Established Species

All seven of the introduced and established natural enemies of the alfalfa weevil in Eastern North America are internal hymenopterous parasites of the egg, larva, or adult stage of the weevil. A brief account is given of the colonization, recovery, biology, and habits of each species. Unless otherwise noted, life-cycle data refer to the Middle Atlantic States. Parasitism figures cited are yearly averages for several locations and fields and are percentages of the *total*, not the peak, host population.

Patasson luna (Girault) (Mymaridae)

This species previously was placed in the genera *Mymar* and *Anaphoidea* and occasionally has been confused with *Anaphes pratensis* Foerster. *P. luna* was imported from Italy and released in Utah in 1911–13 and 1925–28. Material from France was also released in California in 1933–34. In all these introductions, *P. luna* apparently was mixed with *A. pratensis*. The latter was first recovered in 1926 in Utah, but *P. luna* was never recovered in the Western States until recently.³

In 1928, A. pratensis was shipped from Utah to Indiana for release against Hypera nigrirostris (F.), and P. luna might have been included in that material. Although there is no record of a release of P. luna in Eastern North America, in 1958 it was reared from alfalfa weevil eggs collected in Dutchess County, N.Y., by G. O. Poinar of Cornell University. Since then P. luna has been found at about 50 localities in the East (map 1) and is certainly more widely established than is now known.

The adult *P. luna* wasp is 1 mm long. The female deposits one to two eggs inside a weevil egg and may attack several eggs in a cluster within an alfalfa stem. There are several generations each year, and the parasite passes the winter inside the host egg. *P. luna* may have other hosts besides the alfalfa weevil. The data available suggest that parasitism by this species averages from 0 to 10 percent in overwintering eggs of the alfalfa weevil.

Bathyplectes anurus (Thomson) (Ichneumonidae)

This species has also been referred to as *B. anura* and *B. corvina*. Specimens from Europe were released beginning in 1960, and the species was first recovered in New Jersey and Pennsylvania in 1964. The bulk of the introduced *B. anurus* was of French origin, although small numbers came from Sicily and the U.S.S.R. By 1966, field populations of *B. anurus* had increased sufficiently at certain release sites in New Jersey and Pennsylvania to permit collections for recolonization in other States. This work is continuing because the natural dispersion of *B. anurus* is slow. It is now well established in 17 Eastern States and in Ontario. Releases and recoveries of *B. anurus* are shown in maps 2 and 3.

The adult wasp is 3 mm long and has a black, robust body. There is one generation each year. The peak of parasitism occurs about a week before the peak in weevil larvae. The female parasite deposits an egg in the host larva; early instars are preferred. The egg hatches and the parasite larva feeds on the host, killing it after the weevil completes its cocoon. The parasite then spins a hard, brown, football-shaped cocoon, 2 mm in diameter by 3.5 mm long, which bears a raised, white, equatorial band. After formation of the cocoon, the parasite larva enters diapause.

The *B. anurus* larva has the unusual ability of causing its cocoon to "jump" several centimeters upward, a trait shared by few other ichneumonid parasites. This mobility increases the survival of *B. anurus* by enabling many cocoons to escape hyperparasites and unfavorable field conditions. The jumping cocoons can be found in plant debris in alfalfa fields.

In the fall the parasite larva pupates, becomes an adult, and diapauses within the cocoon until early the following spring.

In recent years from 6 to 37 percent of weevil larvae have been parasitized by this species in southeastern Pennsylvania and in central and northern New Jersey.

Bathyplectes curculionis (Thomson) (Ichneumonidae)

This parasite was successfully introduced from

³ P. luna was reared from alfalfa weevil eggs collected in Cache County, Utah, in 1973 (D. W. Davis, Utah State University, pers. commun.).

Italy into Utah in 1911–13 and is now widely distributed in the Western States. In the Eastern United States, releases of B. curculionis from Utah were first made in 1953-55, but these efforts were not successful. In 1959, parasite material obtained from southern California was liberated in Delaware, New Jersey, and Virginia, and in 1960 the species was found to be established in these areas. Although the parental stock of the eastern B. curculionis population is predominantly of Italian origin via California, additional introductions have been made from France, Sweden, and the U.S.S.R. Since its establishment in the East, B. curculionis has been widely recolonized by biocontrol workers (map 4), but it has also spread naturally at a phenomenal rate.

It seems certain that this parasite accompanied the alfalfa weevil as it spread from the eastern seaboard into the Midwest and into Canada. *B. curculionis* has been recorded from more than 750 counties in the East (map 5), and it is probably in every area infested by the eastern strain of the weevil.

B. curculionis adults closely resemble those of B. anurus, but the female of B. curculionis has a longer ovipositor. There is one full generation in the spring, with a peak 1–2 weeks before the host peak, followed by a partial second generation 1–3 weeks after the host peak. Parasite adults are often collected in the summer and fall, especially in the Midwestern States, and these may be the result of a delayed second generation or a partial third or fourth generation.

The female parasite deposits an egg inside the weevil larva, preferring the early instars. The egg may be encapsulated by the host larva's defense reaction, but parasitism is often successful if superparasitism occurs or if the host larva is small when parasitized. Encapsulation of B. curculionis eggs by the host often seriously reduces the effectiveness of this parasite. None of the other established parasites are significantly affected. The surviving parasite egg hatches, the larva develops, and it eventually kills the host. The parasite larva then constructs its cocoon inside the weevil's cocoon like B. anurus. The cocoons of B. curculionis and B. anurus are similar in size and shape, but the white equatorial band of the former is not raised, and the cocoon may be either light brown and flexible (nondiapausing) or dark brown and hard (diapausing). The parasite larva overwinters in the cocoon in the surface litter of alfalfa fields. In the past few years from 5 to 25 percent of weevil larvae have been successfully parasitized by *B. curculionis* in the Middle Atlantic States.

Bathyplectes stenostigma (Thomson) (Ichneumonidae)

This species has been referred to as *Bathyplectes* sp. "bagged," because the cocoon resembles a brown paper bag. It has also been referred to as *B. contracta*. Almost all the *B. stenostigma* released in North America were of Swedish origin; a few lots were from France. Liberations began in 1964, and large numbers were released at many localities in 1968 and 1969 (map 6). The species was first recovered in 1970, but it has been difficult to establish and has dispersed little. To date it has been recovered at only seven release sites in the East (map 7).

The adult parasite resembles the two previous species of *Bathyplectes*, except it is predominantly dark brown and the body is less robust. There is one generation per year, with the peak of parasitism occurring 1–2 weeks after the host larval peak. The female parasite lays an egg in a weevil larva. After hatching, the parasite larva develops, eventually killing the host before the latter spins a cocoon. Thus, the *B. stenostigma* cocoon, which is 3.8 mm long with no external white band, is constructed in the soil litter, where the parasite larva spends the winter in diapause. In a field near Hadley, Mass., during 1974 and 6 years after release, *B. stenostigma* parasitized over 7 percent of the total population of weevil larvae.

Tetrastichus incertus (Ratzeburg) (Eulophidae)

A few authors have called this species *T. erdoesi*. Releases of *T. incertus* began in 1960, and in 1962 it was found to be established in parts of Delaware, Maryland, New Jersey, and Pennsylvania. By 1964, field populations of *T. incertus* were being used as sources of recolonization in other States. Releases of this species are shown in map 8.

The established parasites were from a single locality in southern France; however, from 1967 to 1973, additional small introductions were made from Sweden, Romania, and Yugoslavia to enlarge the gene pool. The parasite dispersed very rapidly during the 1960's, but its movement has

greatly decreased in recent years. It is now present in 16 Eastern States and in Ontario and Quebec (map 9).

The adult is 1–1.5 mm long, black, compact rather than narrow and wasplike. There are three to four generations per year, with the largest populations in July-September. The female oviposits while astride a weevil larva, laying several eggs inside. From four to seven of the resulting parasite larvae complete development, killing the weevil larva after it has spun its cocoon. The parasites pupate within the empty skin of the host, which turns a mahogany brown and becomes brittle after the death of the host larva. Winter is spent in the prepupal stage within the host "mummy."

During July, August, and September of 1965–69, *T. incertus* parasitized from 50 to 80 percent of weevil larvae in New Jersey and the surrounding States, but in recent years the species has been much less abundant. This is a direct result of the much lower levels of alfalfa weevil larvae, especially during the summer and fall. This in turn is a result of the pressure exerted by this and the other parasite species that are abundant in the Middle Atlantic region.

Microctonus colesi Drea (Braconidae)

Before it was named in 1968, this species was eferred to as *Microctonus* sp. "black" or "domestic black." It was first found in 1962 in southeastern Pennsylvania, although it had never been released. Since there is no evidence to suggest that *M. colesi* is native or had gained entrance prior to the alfalfa weevil, it is treated here as an introduced parasite. Probably it entered this country within the weevil. Because *M. colesi* is parthenogenic, establishment could have resulted from a small number of individuals. *M. colesi* is now known to be present in 13 Eastern States and appears to be moving slowly westward, considerably behind the weevil. Releases and recoveries of *M. colesi* are shown in maps 10 and 11.

The adult *M. colesi* is wasplike, 3 mm long, and entirely black. All individuals are female, and there is one generation per year. The period of oviposition roughly coincides with the larval peak of the alfalfa weevil. The egg is laid inside the weevil larva; the larger instars are preferred. After hatching, the first-instar larva enters dia-

pause and remains so while the host pupates and becomes an adult. The parasite larva overwinters within the adult weevil. During the following spring the parasite larva matures, kills the weevil, and spins a white cocoon in the soil litter. Since the fertility of weevils containing $M.\ colesi$ larvae is much reduced in male weevils or eliminated in females, this effect is in addition to the ultimate mortality. From 1969 to 1973, $M.\ colesi$ parasitized from 1 to 18 percent of the "new" (newly emerged, sexually immature) weevils in the Middle Atlantic region.

Microctonus aethiopoides Loan (Braconidae)

For many years this parasite has been referred to as M. aethiops; in fact, this name has been used in practically all the North American literature. Recently, however, Loan decided that the name M. aethiops had been misapplied, and he described the parasite discussed here as M. aethiopoides.

This species was first released against the alfalfa weevil in 1957 and was found to be established in 1961 in New Jersey. All introduced parasites were from France. In 1963, field collections were begun in New Jersey for shipment to and release in other States. This recolonization has continued through 13 seasons. As a result of both recolonization and natural dispersal, *M. aethiopoides* is now present in 14 Eastern States and in Ontario (maps 12 and 13).

The adults are wasplike and 3 mm long; females are red brown and males are black. There are two generations per year. In the spring, about the time of the host larval peak, the female lays a single egg inside many of the "old," overwintered sexually mature weevils. After hatching, the M. aethiopoides larva matures, issues from the dying host, and forms a white, silky cocoon, 4 mm long, in the soil litter. After pupation for 2-3 weeks, the adult parasite emerges from the cocoon. The female parasite oviposits in a "new" sexually immature adult weevil, and the resulting parasite larva diapauses and overwinters in the first instar within the weevil adult. Early the next spring the larvae of M. aethiopoides complete their development, spin cocoons, and transform to adults.

M. aethiopoides kills from 70 to 90 percent of the overwintered weevils in the Middle Atlantic

⁴ Loan, C. C. A review of haliday species of microctonus (hym.: braconidae, euphorinae). Entomophaga 20: 31–41. 1975.

States, and the second generation parasitizes an average of 7 to 39 percent of the new weevils. Even though the parasitized new weevils are not killed until the following spring, all the females and many of the males are rendered sterile, so the control impact is even more significant. Many biocontrol workers in the East consider *M. aethiopoides* the most important single species in the weevil parasite complex.

Species Released but not Recovered

Five of the species discussed here are small Hymenoptera and one is a tachinid fly. These insects were released in Eastern North America during 1957–72 but were never recovered. Possibly a few species are established in certain areas, because many releases have not been followed by surveys adequate for detection. It is hoped that the information provided here, plus the tables and maps, will encourage further searching for these species.

Peridesmia discus (Walker) (Pteromalidae)

This species, previously called *P. phytonomi*, was obtained in southern France and was released from 1959 through 1972. More than 45,000 adults were liberated in the East, mainly in the Southern States (map 14).

The adult is a black, compact wasp, 2 mm long. The female punctures the alfalfa stem with her ovipositor and lays an egg on the weevil egg mass. The larva feeds externally as a predator on the host eggs. Pupation occurs within the stem cavity. There are several generations per year, but because few weevil eggs are present in the summer and early fall, it is likely that *P. discus* passes this period in diapause or on an alternate host. The adults can live for many months and are very hardy. In southern France the adults oviposit in the field from November through January. Extensive surveys in France indicated that this predator was present in 6 percent of the overwintering egg masses.

Trichomalus inops (Walker) (Pteromalidae)

Ten releases were made of this species in the East from 1959 to 1970 (map 15). It was collected in France along with *P. discus*, but it was much less abundant. Both species are similar in size,

appearance, and life cycle as far as is known. In Europe T. inops is also thought to attack the eggs of Apion sp.

Dibrachoides dynastes (Foerster) (Pteromalidae)

This species has also been referred to as *D. druso*. From 1959 to 1964, over 13,000 *D. dynastes* adults of French origin were released at 28 localities in the Eastern States (map 16). The bulk of the released material was reared in the Moorestown laboratory. Although it has never been recovered in the East, in 1967 it was established in San Diego County, Calif., by entomologists of that State.

D. dynastes is a parasite of prepupae and pupae of the alfalfa weevil. The adult parasites are 2–3 mm long, with a metallic-green head and thorax. There are two generations per year in southern France, and the parasite apparently overwinters as an adult. Several eggs are laid on the host prepupa or pupa within its cocoon. After hatching, the parasite larvae feed externally on the body of the host. Usually three to five parasites reach maturity. In Europe this species also attacks other species of Hypera.

Necremnus leucarthros (Nees) (Eulophidae)

This species was released in 1965 at only two localities in New Jersey (map 17). About 300 adults of Italian origin were liberated at each site.

Like the preceding species, N. leucarthros is a gregarious ectoparasite of weevil prepupae. The adult parasites are 1.5–2.3 mm long and metallic green. The males possess branched antennae. In Europe N. leucarthros is widely distributed and attacks several species of Coleoptera, including other Hypera.

Campogaster exigua (Meigen) (Tachinidae)

During 1957, three small releases of this fly were made in Delaware and New Jersey (map 18). A total of 168 adults from France were liberated.

The adults are about 2.5 mm long and dark gray. The female does not lay eggs but deposits a first-instar larva on the thorax of the adult weevil. After penetrating the host, the parasite larva develops and finally forms a brownish puparium within the body of the weevil. The number of generations per year is not known; however, in the laboratory one complete generation requires

from 48 to 70 days. In Europe C. exigua also attacks several species of Sitona.

Microctonus stelleri Loan (Braconidae)

Before it was named in 1972, this species was referred to as *Microctonus* sp. "European black." From 1968 to 1970, over 4,100 adults obtained in France were released at 23 localities in the Eastern States (map 19).

The parasite has one generation per year, and its only known host is the alfalfa weevil. The females of M. stelleri and M. colesi are very similar in size and appearance. However, M. colesi is unisexual (all individuals are female), whereas M. stelleri has both sexes in about equal proportion. In the spring the mature parasite larva issues from the overwintered adult weevil and forms a white cocoon about 4 mm long in the ground litter. In about 2 weeks the adult parasite emerges, mates, and oviposits in larvae of the alfalfa weevil. It prefers larger instars. The parasite egg hatches, the first-instar larva enters diapause, and it remains so while the host pupates and becomes an adult. The M. stelleri larva overwinters within the adult weevil. Extensive collections in northern France during 1966-70 indicated that about 5.6 percent of the overwintering weevils were parasitized by M. stelleri.

Other Parasitic Organisms

Primary Parasites

Included here are various natural enemies that have been recorded from the eastern strain of the alfalfa weevil. These species are native to North America, and most of them attack a variety of other hosts. On the alfalfa weevil their control impact is almost always of minor significance. No doubt additional adventitious parasite species will be discovered.

Fidiobia rugosifrons Crawford (Platygasteridae).—In 1972, adults of this species were reared from overwintering weevil eggs in Ontario. Curiously the parasite has not been recorded from weevil eggs at any other eastern site, although its type locality is in Pennsylvania.

Hexamermis arvalis Poinar and Gyrisco (Mermithidae).—This nematode is most often found in fourth-instar weevil larvae, but pupae

and adults are also parasitized. A single weevil may contain from one to three nematodes. When mature, these nematodes are often more than 9 cm long. The weevil larvae die soon after issuance of the nematodes. This species has been reported from several other insect families.

Several adventitious parasites have been reared from weevil pupae at a few localities in the East: Eriplanus micator (Gravenhorst) (Ichneumonidae), Helicobia rapax (Walker) (Sarcophagidae), Itoplectis conquisitor (Say) (Ichneumonidae), Pediobius sp. (Eulophidae), and Spilochalcis albifrons (Walsh) (Chalcididae). The last species is usually a hyperparasite, attacking Bathyplectes cocoons. See discussion of secondary parasites.

Hyalomyodes triangulifer (Loew) (Tachinidae).—This fly is 3.5 mm long and has been reared from adult weevils collected in the field during the fall, winter, and spring. It is widely distributed in the East and parasitizes many species of adult Coleoptera. It has been recorded from the alfalfa weevil from New York to Georgia. After it issues from the dead host, the fly larva forms a small, brown puparium on the soil. The life cycle is probably similar to that of Microctonus aethiopoides. In certain eastern localities H. triangulifer has occasionally been found in as many as 5 percent of the weevils.

Leucostoma simplex (Fallén) (Tachinidae).— This fly was reared from an adult weevil collected in Dutchess County, N.Y. L. simplex is apparently Holarctic in distribution.

In addition, several diseases attack the alfalfa weevil. Microsporidian diseases, caused by species of *Nosema* and *Perezia*, are common in laboratory cultures, but they are rarely found in the field. Fungus diseases are also known. Those caused by species of *Beauveria* and *Entomophthora* are most frequently observed in nature. Epizootics caused by *E. phytonomi* Arthur have been reported in Ontario during the past few years.

Secondary Parasites

Members of this group, commonly known as hyperparasites, are not parasitic on the alfalfa weevil, but they attack the natural enemies of the weevil instead. They are discussed here only because they will be encountered in parasite impact studies and may be confused with the primary parasites. Competent identification is, therefore, necessary in such investigations.

Because *Bathyplectes* spp. spend many months in cocoons exposed on the soil, they are especially vulnerable to attack by secondary parasites. The following nine hymenopterous species have been reared from cocoons of *Bathyplectes* spp. in Eastern North America. This list does not include a few species that have been reared infrequently from *Bathyplectes*, and certainly additional hyperparasites will be recorded if there are more intensive field investigations.

Spilochalcis albifrons (Walsh)
Eupelmella vesicularis (Retzius)
Gelis sp.
Mesochorus agilis Cresson
Catolaccus aeneoviridis (Girault)
Dibrachys cavus (Walker)
Eupteromalus viridescens (Walsh)
Sceptrothelys grandiclava (Walker
Sceptrothelys intermedia Graham

All these species, except M. agilis, lay their eggs inside the Bathyplectes cocoon; M. agilis oviposits in weevil larvae previously parasitized by Bathyplectes. S. albifrons, E. vesicularis, Gelis

sp., M. agilis, and C. aeneoviridis are solitary parasites, i.e., only one adult is produced per Bathyplectes cocoon. D. cavus, E. viridescens, S. grandiclava, and S. intermedia are normally gregarious. Some unusual morphological features should be noted. Adults of Gelis sp. are always female and wingless, and females of E. vesicularis have greatly abbreviated wings. Adults of S. albifrons have conspicuously enlarged hind femora.

The mummies of Tetrastichus incertus remain on the soil for many months and, like cocoons of Bathuplectes, are particularly exposed to attack by hyperparasites. However, no secondaries are known to have been recorded from T. incertus mummies in this country. Similarly no hyperparasites have been recorded from cocoons of Microctonus, perhaps because they are in the soil litter for only a few weeks. However, the absence of hyperparasite records for these mummies and cocoons may only reflect the limited sampling efforts. On the other hand, we seriously doubt that hyperparasites will be found on the larval stages of Microctonus and Tetrastichus within their hosts. since none have ever been detected in extensive dissections and rearings over a 10-year period at this laboratory.

METHODS FOR RELEASE AND RECOVERY

Making Parasite Releases

Several requisites are critical for the successful establishment of parasites. The alfalfa field selected should have at least a moderate population of alfalfa weevils; the stage of the host should be appropriate (e.g., larvae for T. incertus); and insecticides should not be applied for at least 4 weeks or 1 year or more when possible. Previously mated females or mixtures of male and female parasites are suitable for release. The carton containing the parasites should be placed at the base of the alfalfa plants near the center of the field and opened slightly. If foliage is then arranged over the opening, the parasites will be less likely to disperse widely. After about 5 minutes the container can be examined, and the number and sex of the parasites that died in transit can be determined. It is imperative that parasites be released in the field as soon as possible after receipt.

After a parasite has become well established

at a release site, workers in some States have made recolonization releases within their States to accelerate parasite dispersal. Either parasitized hosts are collected and released directly in a different county, or the adult parasites are first obtained through rearing and then released. In certain States parasite recolonization efforts have been very successful.

Sampling for Recovery

We have found that the number of parasite species attacking the alfalfa weevil in a given area is frequently much greater than had been suspected. Careful surveys are required to determine whether a species has become established at a release site or whether parasites have dispersed from another area. Such surveys are based on (1) field collection of adult parasites, (2) dissection of field-collected hosts, or (3) rearing of parasites from field-collected hosts. Although the first two

methods are extremely useful, they require considerable experience on the part of the collector; they will not be discussed here.

The rearing method begins by collecting the appropriate host stage in the field at the right time. These hosts are placed in the proper type of cage in the laboratory and are provided with adequate food. This should result in parasite adults, cocoons, or mummies, which can then be identified.

Timing of Collections

Egg parasites.—All parasite species that feed internally in the egg or externally on the egg mass can be recovered whenever weevil eggs are present in the field, but the most profitable period is probably during the winter. Alfalfa stems containing weevil eggs should be cut in the field and taken to the laboratory for examination.

Larval parasites.—To provide estimates on timing that will be usable in different climatic regions, it is necessary to relate these estimates to a common biological reference point. The time of the host larval peak or maximum population density in the spring is used here for this purpose. In order to recover the following parasites, host larvae should be collected throughout the periods indicated: Bathyplectes anurus, 14 days prior to the host larval peak; B. curculionis, 14 days prior to and 14 days following the larval peak; B. stenostigma, from the larval peak to 21 days following the peak; Tetrastichus incertus, from the larval peak through the summer and fall if larvae are present.

Pupal parasites.—All species that attack the pupal stage of the alfalfa weevil can be recovered by collecting host cocoons whenever they are most abundant in the field. Depending on locality, this should be about 1–3 weeks after the host larval peak.

Adult parasites.—To recover any of the three species of Microctonus, host adults can be collected during midwinter or when they first appear in the spring. Probably the most profitable time to recover M. aethiopoides from overwintered host adults is from the host larval peak to 21 days thereafter. Hyalomyodes triangulifer can be recovered at this time and also later in the season. In addition, certain workers have successfully used topical applications of synthetic hormones,

such as epoxyfarnesenic acid compounds, on new weevil adults to force emergence of parasites.

Rearing Cages

Egg parasites.—Alfalfa stems can be split to locate weevil egg masses. Eggs can be either left in place in short sections of stem or removed and incubated in small vials, capsules, or petri dishes. Moist filter paper or plaster-of-paris will maintain humidity. Recoveries also can be made by incubating bulk collections of alfalfa stems in darkened emergence chambers and collecting all adult insects attracted to a light source.

Larval parasites.—When attempting recovery of parasites, it is best to collect fourth-instar weevil larvae from the field. Such larvae will have been exposed to parasite attack for the longest time and will require a shorter rearing period in the laboratory. They can be reared in any plastic, wood, or cardboard cage if it (1) is ventilated with two screened side openings, (2) is constructed to prevent escape of the hosts and any parasites that emerge, and (3) contains fresh alfalfa bouquets as food. Another useful technique is to place larvae in large paper bags (up to 2,500 larvae per bag) with a quantity of alfalfa foliage. The bags are cut to a height of 25 cm and the tops left open for ventilation. Weevil larvae remain below with the foliage and do not escape from the bag. Small amounts of fresh alfalfa are added twice each day until the weevil larvae begin to spin cocoons. Then the tops of the bags are closed securely, and the bags are stored in a dry, well-ventilated area for several weeks, after which the parasite cocoons can be removed and counted.

Pupal parasites.—Species that emerge from the pupal stage can be recovered by collecting intact weevil cocoons from ground litter and the lower foliage in alfalfa fields. The host cocoons should be placed individually in small vials or gelatin capsules. By collecting and isolating weevil cocoons in this manner, investigators will obtain not only pupal parasites but many primary larval parasites and their secondaries as well. However, Bathyplectes stenostigma cannot be recovered by this method because it normally kills the host larva before the latter spins a cocoon.

Adult parasites.—To rear parasites from the adult weevil, best results are obtained when weevils are fed alfalfa bouquets in a small, venti-

lated cage with a false screen bottom of about 16 mesh per inch. The parasite larvae will issue from the weevils, pass through the screen, and pupate on the cage bottom under felt strips placed on absorbent paper. Parasite cocoons or puparia should be removed weekly without tearing the fragile cocoons and held in vials for emergence

of adults. Since the larva of Campogaster exigua forms its puparium within the body of the adult weevil, examination of the dead weevils in the cage would be necessary to detect it.

Parasites can be identified by sending the adults to the Systematic Entomology Laboratory, ARS, USDA, ARC-West, Beltsville, Md. 20705.

A RECORD OF PARASITE LIBERATIONS AND RECOVERIES

The seven parasite species presently established in Eastern North America are listed in table 1. Parasite release and recovery information is tabulated alphabetically by species, State, and county, and the locality or the name of the nearest town is given. The year of first recovery in a county is shown in the last column. Many of the releases in table 1 are recolonization releases made by various State workers. Where a parasite has been recovered in a particular State, release and recovery data are presented only on a county basis for that State. It is assumed that biocontrol workers will be able to find specific parasite populations within these counties for purposes of recolonization.

Many individual species have been recovered in counties where the parasite was never released. These records are, of course, the result of natural dispersion. Recoveries in a given field during the same season of release are not included here unless the recovery was made prior to the release. Therefore, the recovery records indicate that a species has survived at least one winter in the field.

The six species released in Eastern North America but not recovered are listed in table 2. As is often the case, the reasons for the failure of these parasites to become established are not known. Unfortunately, attempts to recover introduced species have varied greatly. In a few instances it is still too early to determine whether a species is successfully established or not.

The information on parasite liberations and recoveries has been collected from several sources, including release records on file at the Beneficial Insects Research Laboratory, release and recovery work by our laboratory staff, publications and proceedings of meetings and conferences, and inquiries to individual Federal and State workers. (See cooperators listed under Acknowledgment.)

Certain release information in tables 1 and 2 differs from that in Production Research Report 101, and the present data should be used.

A partial list of parasite releases in the Western States is in table 3 (Appendix).

Table 1.—Species released and recovered in Eastern North America, 1957-75

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Bathyplectes anurus				
Alabama	Limestone	Belle Mina	70	
Delaware	Kent		67, 70	75
	New Castle		65, 67, 69	69
	Sussex		69, 70	70
Illinois		Urbana		
	Pope	Robbs	67	
	Wabash	Kennsburg	68	

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Bathyplectes anurus— Continued				
ndiana	_ Crawford			71
	Floyd			71
	Harrison		65	70
	Lawrence			71
•	Tippecanoe		67	
	Washington			73
owa	Davis	Bloomfield	75	
	do	Pulaski	75	
	Ringgold	Diagonal	75	
Kansas	Geary	Ogden	75	
	Pottawatomie	Flush	74, 75	
	Reno	Hutchinson	74	
	Riley	Manhattan	74, 75	
Kentucky	Caldwell		74	
	Christian		71	
	Fayette		67	70
	Oldham		74	
Maryland	Allegany			75
	Baltimore			75
	Caroline		- -	75
	Carroll		-	75
	Cecil		- -	73
	Frederick			75
	Harford			73
			67	
	Kent			75
	Prince Georges			72
	_			75
	Somerset			75
Massachusetts				74
Michigan				
	Cass		69, 70	
	Ingham		69	70
	-			
			•	
Missouri				71
	Miller		71	
	•			
Vebraska		Gothenburg		
		Lexington		
		Lincoln		
		Gothenburg		
	Otne	Dunbar	75	
		Paul		
		Mead		
		wieau		73

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
New Jersey	Burlington			64
	Camden			
	Cumberland			75
	Gloucester			
	Hunterdon Mercer			
	Middlesex			
	Monmouth			69
	Ocean		04, 00	. 75
	Salem		65, 70, 71	72
	Somerset			69
	Sussex			. 73
	Warren		71	72
New York	Cayuga			70
	Dutchess			. 75
	Franklin		74	
	Herkimer		71	72
	Orange			. 73
	Tomkins		69	70
	Wayne		70	
North Carolina	Ashe		71	
	Rowan			72
	Wake			72
Ohio	Brown		72	
	Clark			
	Clinton			75
	Erie		72	
	Franklin		65	
	Henry Mahoning		75 74	
	Pickaway			
	Putnam			
	Warren			72
	Wayne			. –
	Wood		75	-
Oklahoma	Grady			75
	Kiowa			
	Payne		72, 75	73
	Stephens		72	73
	Washita		75	
Pennsylvania	Berks			67
	Carbon			- 73
	Chester			
	Columbia			- 73
	Cumberland			
	Dauphin			. 71
	Delaware			
	Franklin			. 75

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Bathyplectes anurus— Continued Pennsylvania— Continued				
	Huntingdon			75
	_			64
	•			71
				73
outh Carolina				
				72
ennessee		Newport		
		Crossville		
	Greene	Greeneville	71	
		Knoxville		
	Marshall	Cornersville	71	
		Vonore		
'exas	Burleson	Snook	71, 72	
	Kaufman	Peeltown	71	
	do	Rosser	72	
	Maverick	Eagle Pass	71, 72	
		Iago		
		Lane City		
		Vernon		
·		Crystal City		
rginia				
				71
	Brunswick		71 71	
				73
	Buckingham		72	10
				75

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
	Craig		72	
	Culpeper		71, 73	
	Dinwiddie		71	71
	Fairfax		72	
	Fauquier		71	
	Floyd		72	
	Fluvanna		72	
	Franklin		72	
	Frederick		72	
	Giles		72	
	Gloucester		72	
	Goochland		71	71
	Grayson		72	
	Greene		72	
	Halifax		71, 72	
	Hanover		70, 71, 72	71
	Henrico		72	
	Henry		72	
	Highland		72	
	Isle of Wight		72	
	James City		72	
	King George		72	
	King William		72	
	Lancaster		72	
	Lee		72	
	Loudoun		72	
	Louisa		71	
	Lunenburg		71	
	Madison		72	
	Mecklenburg		72	
	Middlesex		72	
	Montgomery		67, 72	68
	Newport News		72	
	Norfolk		72	
	Northampton		72, 73	
	Nottoway		71	71
			71	
	Pittsylvania		72	
				71
	Prince George		71	71
	Pulaski		69, 71, 73	70

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Bathyplectes anurus— Continued Virginia— Continued				
	Dannahannaala		71	
	Roanoke		72	
	Smyth		72	
	Southampton		71, 72, 73	
	Spotsylvania		71	
Wast Vissisis				
West Virginia				
				72
	-			
				72
			•	
				72
	Monongalia		73	
	Morgan		74	
				71
Wisconsin		_ Leeds		
		Mt. Horeb		
	Vanagha	- 01080	74	
		_ Erly _ Beloit		
Canada (Ontario)				71
Canada (Ontalio)				
Bathyplectes curculionis ²				
Bathyplectes stenostigma ³	N	N 1	90	4.50
Delaware	New Castle	Newark	69	4 70

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
	Sussex	Milton	69	
Illinois	Champaign	Urbana	69	
Indiana	Kosciusko	Pierceton	68	
	Tippecanoe	Lafayette	68	
Kentucky		Lexington		
Maryland	Frederick	Frederick	68	
Massachusetts	•	•		71
Michigan	Berrien	Three Oaks	69	
		Howard T		
	do	Milton T	68	
		Pokagon T		
		Gull Lake		
New Hampshire				70
New Jersey	-			
		Rancocas		
		Alloway		
		Griggstown		
		Newton		
		Washington		
New York		Mohawk		71
		Dansville		
		Sheds		
	-	Ithaca		72
0.14		S. Lansing		
Ohio		S. Bloomfield		
		Wooster		4 72
D 1		Bowling Green		
Pennsylvania		Wysox		
		Sylvan		
		Bowmansville		
		Christiana		
		Martins Creek		
Vones en t		Brotherton		
Vermont		Shoreham		
Vincinia		Shelburne		
Virginia		Charlotte C.H.		
Wissersin		Elliston		
Wisconsin		Pleasant Prairie Beloit		
Canada (Ontario)				⁴ 71
Microctonus aethiopoides 5				
Arkansas	Craighood	Monotto	67	
ni naiisas	0	Monette Keiser		
Connecticut		Keiser		69
Delaware				68
				67
T11.		Homer		_ 00
Illinois \dots				

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Microctonus aethiopoides Continued	5			
Illinois— Continued				
	Clark	_ Casey	66	
		Lerna		
		Palestine		
		Neoga		
		Equality		
		Vienna		
		Taylor Springs		
Indiana	_	Dubois		
		New Albany		
	*	Laconia		
		Vincennes		
	La Porte		74	
		Klondike		
	do			
.owa	Lee	-		
Kansas		Ashland		
		Manhattan		
Kentucky	*	N. Middletown	•	
xentucky		Lexington		'1
		Hustonville		·
Maine	Cumberland			
Maryland				
viaryland				71
	Caroline		•	. 70
	Cecil			_ 70
	Frederick			_ 69
	Harford			_ 70
	Howard		70	72
	Kent			_ 70
				70
	~			70
	Prince Georges			
	Somerset			72
				70
Massachusetts				
massachusetts	Berkshire Franklin			_ 70 70
				70
Michigan	Hampshire			
Aichigan	Allegan			- 73 75
	Arenac Barry			
	_			
	Benzie			_ 19
	Berrien			7°
	Branch			75 70
				_ 73
				70
			•	75
	Eaton			_ 73

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
	Hillsdale		73	
	Ingham		70, 73, 74	72
	Ionia		73	75
	Isabella		73	75
	Jackson		69	70
	Kalamazoo		69, 73	70
	Kent		73	
	Lake		74	75
	Lapeer		73	75
	Lenawee		68, 73	75
	Livingston		73, 74	75
	Manistee			75
	Mason		74	75
	Mecosta		73	75
	Missaukee		70, 71, 72	75
	Monroe			75
	Montcalm		73	75
	Muskegon		73	
	Newaygo			75
	Oceana			75
	Osceola		73	75
	Oscoda			_ 75
	Ottawa		73	
	Saginaw		73	75
	St. Clair			
	Sanilac			75
	Shiawassee		•	75
	Tuscola			72
	Washtenaw			75
Minnesota	Houston		*	
Mississippi	Oktibbeha			
Missouri	Boone			
	Cape Girardeau			
	Grundy		71	
Nebraska	Dawson			
	Lincoln	-		
	Otoe			
	Saline			
	Saunders			
New Hampshire	Cheshire			_ 73
F			67	
	Hillsborough			73
	Merrimack		66	
	Strafford		66	68
New Jersey				_ 71
,	Burlington			61
	Camden			_ 69
	Cumberland			
	Gloucester			65
	Hunterdon			61

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Microctonus aethiopoides Continued	5			
New Jersey— Continued				
	Mercer		65	67
	Middlesex			_ 69
	Monmouth			65
				- 69
	Ocean			- 75
	Passaic		65	
	~ .			69
	Somerset		62	64
	Sussex			_ 68
	Warren		62, 65, 67	68
Jew York	Allegany		74	
	Broome			_ 71
	Cattaraugus		72, 74	
	Cayuga		68	70
	Chautauqua		72, 74	
	Chemung			_ 73
	Chenango			
	Clinton			71
	Columbia			_ 70
	Cortland			73
	Delaware			
	Dutchess			_ 70
	Erie			
	Franklin			71
	Fulton			_ 71
	Genesee Herkimer			70
	Jefferson			74
	Lewis			_ 71
	Livingston			72
	Madison		,, ,, ,,	
	Monroe			74
	Montgomery			_ 71
	Niagara			
	Oneida			_ 74
	Onondaga			_ 72
	Ontario		71, 73, 74	74
	Orange			_ 69
	Orleans		74	
	Oswego		70	73
	Otsego			
	Rensselaer			
	St. Lawrence		72, 74	73
	Saratoga			
	Schoharie			
	Schuyler			
	Seneca		71, 72	73

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
	Steuben		71, 74	71
	Tioga		65, 66	70
	Tompkins		68, 71	70
	Ulster		68	69
	Washington			_ 75
	Wayne		73	74
	Wyoming		72, 74	
	Yates		72	73
North Carolina	Rowan	Salisbury	66	
	Wake	Raleigh	63	
)hio	Auglaize			₋ 75
				75
	Clinton		73, 75	74
	Coshocton		70	
	Erie		71	
	Guernsey		70	
	Hancock			_ 75
	_			75
			,	
	•			
				73
	Wood			75
	Wyandot			₋ 75
Oklahoma	Payne	Stillwater	73	
		Duncan		
Pennsylvania				71
_	Berks			- 68
				69
	Bucks			_ 69
	Butler		70	
				69
				68
				- 73
				0
				68
	<u>-</u>			- 75
				71
	77. 1			
				68
				(5)
	Luzerne			
	Monroe			- 69

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Microctonus aethiopoides 5- Continued	_			
Pennsylvania— Continued				
	Northumbarland			79
				10
				71
			· -	73
Rhode Island				73
rennessee		Crossville		
		Knoxville		
	Marshall	Lewisburg	66	
Гехаs		College Station		
Vermont				70
				73
	_			69
	Grand Isle		68	69
	Washington			_ 71
	Windham			_ 73
Virginia	Accomack		72	
	Albemarle		72	
	Alleghany		72	
	Amelia		71, 73	
	Amherst		72	
	Appomattox		72	
	Augusta		72	
	Bath		72	
			•	
	Bland		72	
	_			
				75
	Clarke		72 79	75
	-10yu		. 4	

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year o first recover (19—)
	Franklin		_ 72	
	Frederick		72	
	Giles		- 72	
	Goochland		71	
	Grayson		72	
	Greene		72	
	Halifax		72	
	Hanover		67, 69, 71, 72	68
	Henrico		72	
	-			
	James City		72	
	King and Queen		_ 72	
	King George		- 72	
	Lancaster		72	
	Lee		72	
	Loudoun		- 72	
	Louisa		71	
	Lunenburg		- 71	
	Madison		_ 72	
	Mecklenburg		_ 72	
	Middlesex		_ 72	
	Montgomery		65, 67, 72	73
			_ 71	
	Nelson			
	New Kent		_ 71	71
			_ 72	
	Norfolk		_ 72	
	Northampton		72, 73	
	_			
	Powhatan		,	71
	_			
	Prince William		_ 72	
				72
	-			
			. / */	

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Microctonus aethiopoides 5— Continued	-			
Virginia—				
Continued				
	0 1		5 0	
	-			
	• •			
			. –	
	_			
	Westmoreland		. 72	
	Wise		72	
VI t VI' ' '	-		. 71	73
Vest Virginia	Berkeley		70	
T7*		A .124		
Wisconsin		Arlington		
		Mt. Horeb		
		Erly		
		Beloit		
1 (0 : :)		Sauk City		
Canada (Ontario)				
	Dundas		. 71	
				5 0
	_		•	72
	Grenville			
	Haldimand		•	71 72
	Halton		. 71	72
	Hastings		. 71	
	Kent		. 70	71
	Lambton		. 71	
			71	
	Lincoln			
			71	
	T) 1		71	
			71	
				70
				72 79
				72
Microctonus colesi 6				
Connecticut	Tolland			67

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Delaware				
	Sussex			
llinois		Ivesdale		
		Urbana		
		Roslyn	,	
		Clinton		
	Douglas	Camargo	67	
		Hindsboro		
	Madison	Alhambra	68	
	Washington	New Minden	67	
ndiana	_	Thorntown		
	Montgomery	Waynetown	67	
Kentucky	Fayette	Lexington	70	
Maryland	Allegany			70
	Anne Arundel			71
	Baltimore			_ 71
	Carroll			- 67
	Cecil			_ 62
	Dorchester			_ 71
	Frederick			_ 67
	Garrett			_ 71
	Harford			_ 69
	Howard			_ 66
	Montgomery			_ 66
	Prince Georges		68	66
	Queen Annes			_ 71
	Washington			_ 67
Massachusetts				_ 70
Michigan		Three Oaks		
		Howard T		
	Jackson	Springport	69	
New Jersey				
	Cumberland			- 66
				_ 69
				_ 69
	Somerset			
				_ 69
7 77 1				
New York				
				77.0

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Microctonus colesi 6— Continued New York—				
Continued				
	Schuyler			_ 73
	Tompkins			
	Ulster			
North Carolina	Rowan			_ 70
	Wake			_ 67
)hio	Belmont			_ 70
	Brown		72	
	Clark		74	
	Clinton	s5,	74	
	Delaware			_ 75
	Erie			
	Franklin			
	Guernsey			
	Hancock			_ 74
	Logan			
	Meigs			_ 72
	Ottawa			
	Putnam		•	
	Seneca			
	Shelby			
	Union			75
	Warren Washington			71
	Wayne			
	Wood			- 1±
Pennsylvania	Bedford			
	Berks			
	Bradford			
	Bucks			
	Butler			
	Centre			
	Chester			_ 62
	Clarion			_ 71
	Cumberland			_ 71
	Dauphin			_ 71
	Franklin			_ 71
	Huntingdon			_ 72
	Jefferson			_ 71
	Lancaster			_ 65
	Lebanon			
	Luzerne			
	Mercer			
	Monroe			
	Montour			
	Northampton			
	Northumberland			
	Pike			
	Snyder			- 71

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
	Somerset			_ 69
	Union			_ 71
	Washington			_ 72
	Westmoreland			_ 71
	York			
Rhode Island	Washington			=0
Vermont	Bennington			
	Accomack			- 12
Virginia	Albemarle			
			72	
	Alleghany		72	
	Amelia		73	
	Amherst		72	
	Appomattox		72	
	Augusta		72	73
	Bath		72	
	Bedford		72, 73	
	Bland		72	
	Buchanan		72	
	Buckingham		72	
	Campbell			
	Caroline		72	
	Carroll		72	
	Charles City		72	
	Charlotte			_ 73
	Chesterfield		72	
	Clarke		72	70
	Craig		72	
	Culpeper		73	
	Cumberland		72	
	Dickenson		72	
	Dinwiddie		71	71
	Fairfax		72	• 1
	•			
	Fluvanna		72	
	Franklin		72	
	Frederick		72	70
	Giles		72	
	Gloucester		72	
	Grayson		72	
	Greene		72	
	Halifax		72	
	Hanover		71, 72	71
			72	
	King William		72	
	Lancaster		72	
	Loo		79	

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Microctonus colesi ⁶ — Continued irginia—				
Continued				
	Loudoun		_ 72	
	Madison		_ 72	
				68
	*			
				70
	_			
				73
			·	
		-		
Test Vincinia				79
Vest Virginia	-			73
				72
	-			72
				70
				70
7*				
Visconsin				
anada (Ontario)		Pleasant Prairie South Bay		
Patasson luna				
elaware				
	New Castle			60
llinois				
	Hardin			

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Indiana	Harrison			72
Maryland	Cecil			64
	Howard			65
	Prince Georges			62
Massachusetts				63
	Hampshire			64
New Jersey	Burlington			61
	Warren			62
New York				71
	Dutchess	· 		58
	Livingston			73
	Tompkins			71
Ohio	Clinton			73
	Columbiana			66
	Erie			69
	Franklin			73
	Lawrence			66
	Mahoning			66
	Meigs			
	Wayne			69
Pennsylvania				62
	Fulton			75
	Lancaster			
	Philadelphia			64
Virginia				73
West Virginia				71
	Hardy			
	Mason			71
	Preston			68
Canada (Ontario)	Carleton			70
	Elgin			73
	Essex			69
	Haldimand			73
	Halton			73
	Kent			73
	Lambton			73
	Lincoln			73
	Middlesex			73
	Oxford			73
				73
	Prince Edward			72
	Waterloo			73
	Welland			73
a 1 /0 :	Wellington			
Canada (Quebec)	Missisquoi			69
Tetrastichus incertus				
Connecticut	Tolland			64
Delaware	Kent			64
	New Castle			
	Sussex			

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Tetrastichus incertus— Continued				
llinois	Bond	Donnellson	66	
		St. Joseph		
		Urbana		
	Christian	Sicily	67	
	Coles	Fox Ridge State Park	65	
		Kenney		
		Albion		
		Watson	,	
		Vandalia		
	-	Duncan Mills		
		Equality		
		Cave in Rock		
		Carbondale		
		Grimsby	66	
		Ste. Marie	65	
	_	Texico		
		Vienna		
		Mt. Pulaski		
	_	Le Roy		
		Boody		
		Alhambra	66	
		Havana		
		Mermet		
		Atterberry		
		Lodge		
		New Hartford		
		Eddyville		
		· ·		
		Mounds	64	
		Sparta		
		Fayetteville	66	
		Galatia	65	
		Mechanicsburg		
		Winchester		
		East Lynn	67 cc	
diana		Brownsville	66	
iuiana		Dove		
		Jamestown		
		Corydon		
	Jackson	Brownstown	66	
		Samaria		
		Wanatah		
		Shannondale		
		Smartsburg		
		Ballstown		
	do	Osgood	66	
		Manhattan		
entucky		Gracey		
	Favette	Lexington	71	

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
	Lincoln	(?)	64	
	Montgomery			_ 4 65
	Pulaski	(?)	64	
	Todd	(?)	64	
Maine	Cumberland			73
Maryland	Allegany			74
	Anne Arundel			64
	Baltimore			64
	Carroll			64
	Cecil			62
	Frederick.			65
	Garrett			74
	Harford			63
	Howard			64
	Kent			65
	Montgomery			64
	Prince Georges		_ 65	65
	Queen Annes			65
	Washington			65
Massachusetts	Berkshire		_ 67	65
	Bristol		_ 64	66
	Essex			66
	Franklin		_ 62	65
	Hampden			65
	Hampshire		_ 62, 67	65
	Middlesex			67
	Norfolk			_ 66
	Plymouth			_ 66
Michigan	_ Branch			
	Calhoun			
	Cass			
	Eaton		_ 68	72
	Hillsdale			
	Ingham			70
	Jackson			10
			_ 68	72
	Kalamazoo Lenawee			
	Monroe		_ 68, 69	73
	St. Joseph		_ 68	73
	Van Buren			
Mr.	Washtenaw			
Missouri				
	Carter			
	New Madrid			
	Pemiscot			
	Reynolds			
	do	(?)	64	
	Stoddard	(?)	64	
	Texas			
Nebraska	Dawson	Gothenburg	_ 73	

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Tetrastichus incertus— Continued				
New Hampshire	_ Cheshire			65
	Grafton			68
	Hillsborough			70
	Merrimack			68
	Strafford		67	68
	Sullivan		67	
New Jersey				63
	Camden		•	64
	Cumberland		*	69
	Gloucester		· ·	64
	Hunterdon		62, 63	64
	Mercer		62, 63, 64, 65	63
	Middlesex		63, 64, 65	64
	Monmouth		62, 63, 64, 65	64
	Morris		62, 63	63
	Ocean			75
	Salem		63, 64, 65	64
	Somerset		62, 63, 64	64
	Sussex		63	63
	Warren		60, 61, 62, 63	62
New York	Albany			65
	Allegany			66
	Broome			64
	Cattaraugus			66
	Cayuga		65, 68	67
	Chautauqua			66
	Chenango			65
	Clinton			68
	Columbia		63	65
	Cortland			65
	Delaware			64
	Dutchess			65
	Erie			67
	Essex			68
	Franklin			68
	Fulton			66
	Genesee			67
	Greene			65
	Herkimer			67
	Jefferson			
	Lewis			
	Livingston			
	Madison			
	Monroe			
	Montgomery			66
	Niagara			67
	Oneida			67
	Onondaga		CE	66

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
	Orange			_ 63
	Orleans			_ 67
	Oswego			_ 67
	Otsego			_ 65
	Rensselaer			_ 65
	St. Lawrence			_ 69
	Saratoga			_ 66
	Schenectady			68
	Schoharie			_ 67
	Schuyler			_ 65
	Seneca			_ 66 _ 65
	Steuben Sullivan			_ 64
			64 65	65
	Tompkins		. *	66
	Ulster			64
	Warren		00	_ 66
	Washington			0.5
	Wayne			_ 67
	Wyoming			_ 66
	Yates			_ 66
North Carolina	Ashe			_ 72
	Buncombe			_ 74
	Haywood			_ 73
	Orange		61	
	Rowan		71	72
	Wake		61, 65, 71	72
)hio	Allen		72	
	Ashland		67	
	Ashtabula			_ 71
	Butler			71
	Carroll Clark		66 65	
	Columbiana		66	70
	Coshocton		00	_ 71
	Crawford		71, 72	
	Darke			
	Defiance		72	
	Erie		72	
	Fairfield			_ 73
	Fayette		72	
	Franklin			
	Greene		72	
	Guernsey			
	Harrison			
	Huron			
	Jefferson			
	Knox			
	Lorain			
	Lucas		72	

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Tetrastichus incertus— Continued				
Ohio— Continued				
	Medina		65. 67	71
	_			
	_			
				_ 71
	U			70
Oklahoma		Duncan		
Pennsylvania	_			_ 64
				_ 65
				_ 63
				_ 69
				_ 70
	Centre		61	63
	Chester		61	62
	Clarion		62	73
				_ 64
	Clinton			_ 64
				_ 67
	Cumberland			_ 70
				_ 64
	_			_ 64
	Erie			_ 64
	Franklin			_ 64
	Fulton			_ 64
	Huntingdon			_ 64
				70
	Lancaster			_ 62
	Lebanon			_ 70

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
	Perry			- 64
	-			65
	•			
	-			
	-			70
South Carolina		Newberry		00
Γ ennessee		Maryville		
· ciiiicosce		Greeneville		
		Knoxville		
	Madison		63	
		(:) Cornersville		
	Monroe			
Гехаs		Snook		
exas		Rosser	,	
		Eagle Pass		
		Cottondale		
		Iago Vernon		
	Zevele	Crystal City	71 79	
Vermont		Crystar Orty		70
vermont				70
				71
				70
			CE	67
7ingini				01
Virginia				
				70
	Amherst			
	_			68
			-	
				69
				66
	-			
	Chesterfield		72	

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Tetrastichus incertus— Continued Virginia— Continued				
001111111111111111111111111111111111111	Cl. 1		7 0	
	Cumberland		. 72	
	Dickenson		. 72	
	Dinwiddie		. 71	71
	Fairfax		. 72	
	Fauquier			
			•	
	Giles		–	66
	Gloucester		· .:	
			,	71
	Grayson		. 72	
	Greene		. 72	
				72
			67, 71, 72	68
	Henrico		. 72	
	Henry		61, 72	
	Highland		. 72	67
	Isle of Wight		. 72	
	James City		. 72	
	King and Queen		. 72	
	King George		. 72	
	King William		. 72	
	Lancaster		. 72	
	Lee		. 72	
	Loudoun		. 72	
	Louisa		. 71	
	Lunenburg		. 71	
	Madison		. 72	
	Mecklenburg		. 72	
	Middlesex		. 72	
	Montgomery		61, 72	66
	Nansemond		. 71	
	Nelson		. 72	
			69, 71	71
	Newport News		. 72	
	Norfolk		. 72	
	Northampton		72, 73	
	=	-		71
	_			
	_			
	Powhatan			71

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released	County	Locality	Year of release	Year of first
and State			(19—)	recovery (19—)
	Prince Edward		72	
	Prince George		71	71
	Prince William		72	
	Pulaski		71, 73	67
	Rappahannock		71	
	Richmond		72	
	Roanoke		72	
	Rockbridge		72	
	Rockingham		72, 73	
	Russell		72, 73	
	Scott		72	
	Shenandoah		72	65
	Smyth		72	
	Southampton		71, 72, 73	
	-			
	Tazewell		72	
	Washington		72	
	_			
	Wise		72	
Vest Virginia				71
				70
				71
				65
	Mason		_ 71	72
	Mineral			65
	Monroe			
	Preston			70
Visconsin		Arlington		
		Lodi		
	Iowa		73	
	Kenosha	· /	69	
		Pleasant Prairie	69	
	Marquette		73	
	•	(*) Gotham		
Canada (Ontario)	Brant		- 70	72
ranada (Ontario)	Bruce		10	71
				70
				71
	Elgin		70	72
	9			
	Hrontongo			
	Frontenac			
	Grenville			
	Grenville Haldimand			72
	Grenville Haldimand Halton			72 71
	Grenville Haldimand Halton Hastings			- 72 - 71 - 72
	Grenville Haldimand Halton Hastings Kent		70	- 72 - 71 - 72 72
	Grenville Haldimand Halton Hastings Kent Lambton		70 70	- 72 - 71 - 72 - 72
	GrenvilleHaldimandHaltonHastingsKentLambtonLincoln		70 70 70	- 72 - 71 - 72 72

Table 1.—Species released and recovered in Eastern North America, 1957-75—Con.

Species released and State	County	Locality	Year of release (19—)	Year of first recovery (19—)
Tetrastichus incertus—				
Continued				
Canada (Ontario)—				
Continued				
	Northumberland			_ 72
	Peel			_ 73
	Prince Edward		70	72
	Waterloo		70	71
	Welland		70	
	Wellington			_ 71
	Wentworth			_ 71
Canada (Quebec)				

¹ East of longitude 100° W.

Table 2.—Species released in Eastern North America 1 but not recovered, 1957-75

Species released and State	County	Locality	Year of release (19—)
Campogaster exigua ²			
Delaware	New Castle	St. Georges	57
		Moorestown	
	do	Mount Holly	57
Dibrachoides dynastes 3			
Georgia	Clarke	Athens	62
Illinois	Hardin	Cave in Rock	64
	Johnson	Vienna	64
	Williamson	Creal Springs	64
Maryland	Howard	Clarksville	60
Massachusetts	Franklin	Leverett	62
	Hampshire	Amherst	64
	do	Leverett	62
Missouri	Reynolds	(?)	64
	Stoddard	(?)	64
New Jersey	Burlington	Moorestown	64
	do	Mount Holly	59, 60, 63
	do	Rancocas	59, 60
	Hunterdon	Flemington	62
	do	Ringoes	60
See footnotes at end of table			

² Release and recovery counties for *B. curculionis* are shown on maps 4 and 5. This species is now so widely distributed that it is unnecessary to tabulate release and recovery data.

³ Because recoveries of B. stenostigma have been infrequent to date, place localities are shown for all releases.

⁴ Permanent establishment at this locality is in doubt. Subsequent recovery attempts have been negative.

⁵ List includes only releases against alfalfa weevil. During 1948–57, several releases of *M. aethiopoides* were made against *Sitona* spp. in Manitoba, Minnesota, Nebraska, North Dakota, and Washington, but no recoveries are known.

 $^{^6\,}M.\ colesi$ was not purposely introduced, but it is probably of foreign origin. See discussion on page 4.

Table 2.—Species released in Eastern North America ¹ but not recovered, 1957-75
—Continued

Species released and State	County	Locality	Year of release (19—)
	Mercer	Pennington	62
	Somerset	Belle-Mead	62
	Warren	Blairstown	60, 61, 62
	do	Johnsonburg	61
New York	Tioga	Nichols	64
North Carolina	Wake	Raleigh	63
		State College	
•		Christiana	
Virginia	Craig	Newcastle	61
3		Wirtz	
West Virginia			61
	Monongalia		61
	Monroe		61
Microctonus stelleri			
	New Castle	Newark	69
	Sussex	Overbrook	69
Illinois	Champaign	Urbana	69
	Douglas	Kemp	68
		Fairfield	
Kentucky		Kevil	
•		Lexington	69
Maryland		Frederick	
		Northampton	
Michigan	Berrien	Three Oaks	69
		Howard T	
		Pokagon	
New Jersev		Rancocas	
1.0W 00100g		Alloway	
		Hamburg	
New York		New Paltz	
Panneylyania	Franklin	Sylvan Sylvan	69
remisyrvama		Martins Creek	
		Dillsburg	
Vormont		Shoreham Shoreham	
virginia		Steeles Tavern	
Wissessin		Charlotte C.H.	
	_ Rock	Beloit	10
Necremnus leucarthros	70 11 (To a second	0.5
new Jersey		Deacons	
	Mercer	Pennington	65
Peridesmia discus			
		Decatur	70
Arkansas	Craighead	Monette	67
	Mississippi	Blytheville	
	do	Keiser	66
		Yarbro	
Delaware		Dover	
	do	Smyrna	59
	New Castle	Middletown	59
Georgia		Athens	
		Roswell	
		Bogart	

Table 2.—Species released in Eastern North America ¹ but not recovered, 1957-75
—Continued

Species released and State	County	Locality	Year of release (19—)
Peridesmia discus— Continued			
Illinois	Hardin	Cave in Rock	70
Kentucky	Fayette	Lexington	72
Maryland	Howard	Clarksville	60
		Columbus	
	do	Medford	59
	do	Mount Holly	60
North Carolina	Rowan	Salisbury	66
	Wake	Raleigh	66, 67
	do	Wake Forest	67
Oklahoma	Stephens	Duncan	72
Pennsylvania	Chester	Oxford	61
South Carolina	Greenwood	Ninety Six	72
	Oconee	Clemson	72
Tennessee	Cumberland	Crossville	72
	Knox	Knoxville	67
	Marshall	Lewisburg	66
	Maury	Spring Hill	66
	Sullivan	Blountville	72
Texas	Burleson	Snook	71, 72
	Kaufman	Rosser	72
		Eagle Pass	
	do	Quemado	71
		Boling	
	do	Iago	72
	Wilbarger	Vernon	72
	Zavala	Crystal City	71, 72
Virginia	Accomack	Onancock	59
		Red Oak	
	Montgomery	Blacksburg	72
	Rockbridge	Steeles Tavern	72
Trichomalus inops			
Alabama	Morgan	Decatur	70
Arkansas	Mississippi	Keiser	67
Delaware	Kent	Dover	59, 60
	do	Smyrna	59
	New Castle	Middletown	59
		Cave in Rock	
New Jersey	Burlington	Columbus	59
	do	Mount Holly	60
Virginia	Accomack	Onancock	59

¹ East of longitude 100° W.

 $^{^2}$ List includes only releases against alfalfa weevil. During 1948–57, several releases of $C.\ exigua$ were made against Sitona spp. in Manitoba, Minnesota, Nebraska, North Dakota, and Washington, but no recoveries are known.

 $^{^3}$ Although D. dynastes has never been recovered in the East, it is known to be established in California.

APPENDIX

Table 3.—Partial list of species released in the Western States ¹ and their origin, 1957-75

[Includes only parasite material supplied through USDA sources]

	released,		Year of	
State, ar	nd county	Locality	release (19—)	Origin
Bathyplectes anurus			~	
	Alameda	Albany	67	N.J.
	Glenn	Orland	67	N.J.
	Riverside	Lakeview	67	N.J.
Colorado	Garfield	New Castle	75	Pa.
	Larimer	Ft. Collins	75	Pa.
	Mesa	Fruita	75	Pa.
	do	Palisade	71, 73, 74, 75	N.J., Pa.
daho	Latah	Moscow	69	N.J., Pa.
Montana	Ravalli	(?)	71	Pa.
Vevada	Churchill	Fallon	71	Pa.
	Washoe	Reno	71	Pa.
New Mexico	Valencia	Los Lunas	75	Pa.
Oklahoma	Texas	Guymon	74	Pa.
Oregon	Benton	Corvallis	74, 75	Pa.
	Grant	Dayville	72	N.J., Pa.
	Linn	Corvallis	72	N.J., Pa.
South Dakota	Lawrence	Spearfish 2	71, 72	N.J., Pa.
Jtah	Cache	Logan	72	N.J., Pa.
Washington	Clallam	(?)	71	Pa.
Wyoming	Fremont	Riverton	71	Pa.
	Platte	Wheatland	73	N.J.
Bathyplectes stenostigme	7			
	~ Alameda	Albany	69	Sweden.
	Mesa	•		Do.
301014401111111111111111111111111111111		Fruita 3		Do.
Utah	Cache			Do.
		Troitin Boguin	00	20.
Dibrachoides dynastes	3.5	(0)		
Colorado	Mesa	(?)	64	France.
Microctonus aethiopoide	8			
Arizona	Maricopa	Phoenix	65	N.J.
	do	Tempe	65	N.J.
	Yuma	Yuma	65	N.J.
California	Riverside	Crestmore	68	N.J.
	Siskiyou	(?)	66	N.J.
Colorado	Mesa	Clifton	65	N.J.
	do	Collbran	73	N.J.
	do	Grand Junction	67, 69	N.J.
		Palisade		N.J.
	Montezuma	Dolores	73	N.J.
	do	Lewis	73	N.J.
See footnotes at end of table.				

Table 3.—Partial list of species released in the Western States ¹ and their origin, 1957-75—Continued

Species re State, and		Locality	Year of release (19—)	Origin
Microctonus aethiopoides- Continued	_			
daho	Bingham	Aberdeen	66	N.J.
		Juliaetta		N.J.
		Moscow		N.J.
	Twin Falls	Twin Falls	66	N.J.
Iontana	Ravalli	(?)	71	Pa.
Vevada	Washoe	Reno	71	N.J.
lew Mexico				Pa.
regon	Benton	Corvallis	72, 73, 74	N.J., Pa.
		do		N.J.
outh Dakota				N.J.
Jtah				N.J.
	do			N.J.
Vyoming				N.J.
	do			N.J.
	Platte	Wheatland	72	N.J.
Microctonus colesi				
California	Alameda	Albany	69	N.J.
		La Sierra		N.J.
		Dorris		N.J.
	•	Yreka		N.J., Pa.
Colorado	Mesa	Grand Junction	69	N.J.
daho	Latah	Juliaetta	69	N.J.
16:				
Microctonus stelleri	16.	CI:ti	0.0	Th
Colorado		Palisade		France. Do.
	uo	ransade	09	ъо.
Peridesmia discus				
rizona	Maricopa	Tempe	67	Do.
California	Monterey	Gonzales	72	Do.
Vevada				Do.
regon	Linn	Corvallis	72	Do.
Tetrastichus incertus				
rizona	Maricopa	Phoenix	66	N.J.
	do			N.J.
		Yuma		N.J.
California			63	France.
		Hemet	65	N.J., Pa.
		Lakeview		France,
				N.J., P
	do	La Sierra	65	N.J., Pa.
		Riverside		France,
				N.J., P
	do	(?)	66	N.J.
	San Diego	San Pasqual	64, 65	France, N.J., P
	do	(?)	66	N.J.
	Siskiyou	(?)	63, 66	

Table 3.—Partial list of species released in the Western States ¹ and their origin, 1957-75—Continued

	pecies released, ate, and county	Locality	Year of release (19—)	Origin
Colorado	Delta	(?)	64	N.J.
	Garfield	New Castle	64	N.J.
	do	Rifle	64	N.J.
	Larimer	Ft. Collins	65	N.J., Pa.
	do	Kelim	66	N.J., Pa.
	do	Timnath	65	N.J., Pa.
	Mesa	De Beque	64	N.J.
	do	Fruita	65, 71	N.J., Pa.
	do	Grand Junction	69	N.J.
	do	Loma	65	N.J., Pa.
	do	Palisade	64	N.J.
	Montrose	(?)	64	N.J.
	Weld	Berthoud	66	N.J., Pa.
	do	Gilcrest	65	N.J., Pa.
	do	Greeley	65	N.J., Pa.
	do	Johnstown	66	N.J., Pa.
daho	Bingham		65, 69	N.J., Pa.
	Canvon	Caldwell	64	N.J., Pa.
	Caribou	Grace	65	N.J., Pa.
	Latah			N.J.
	do			N.J., Pa.
	Twin Falls	Twin Falls	65	N.J., Pa.
Nevada	Washoe			France.
Oregon	Linn	Corvallis	73	N.J., Sweden
South Dakota	Lawrence	Spearfish	69	N.J., Pa.
	Cache			N.J.
		Draper	67	N.J.
	do			N.J.
Washington	Whitman			N.J.
	Platte			N.J., Sweden

¹ West of longitude 100° W.

² Recovered during 1973.

³ Recovered during 1971 and 1975.

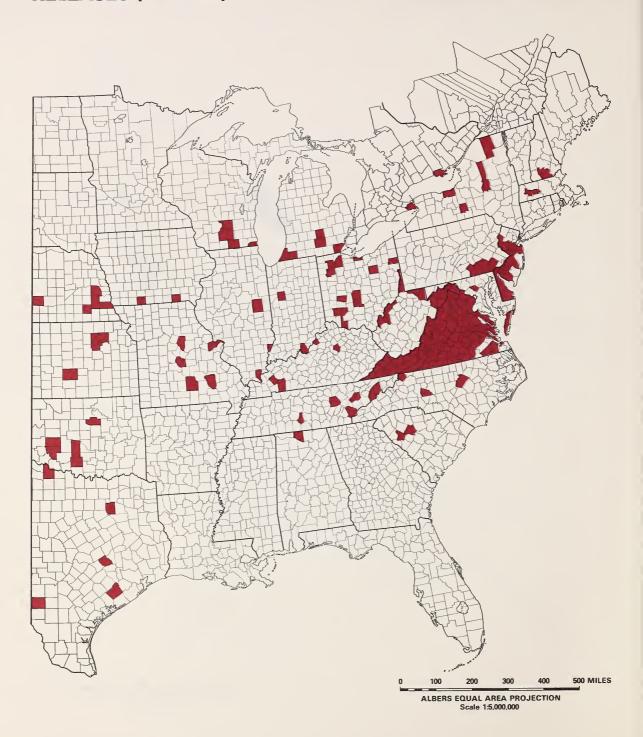
Note.—Many other parasite colonies have been liberated in the West. Since California has had an active parasite introduction program against *Hypera postica* and *H. brunneipennis* for many years, the California reports and publications should be consulted for a complete record.



PATASSON LUNA RECOVERIES (TABLE 1)



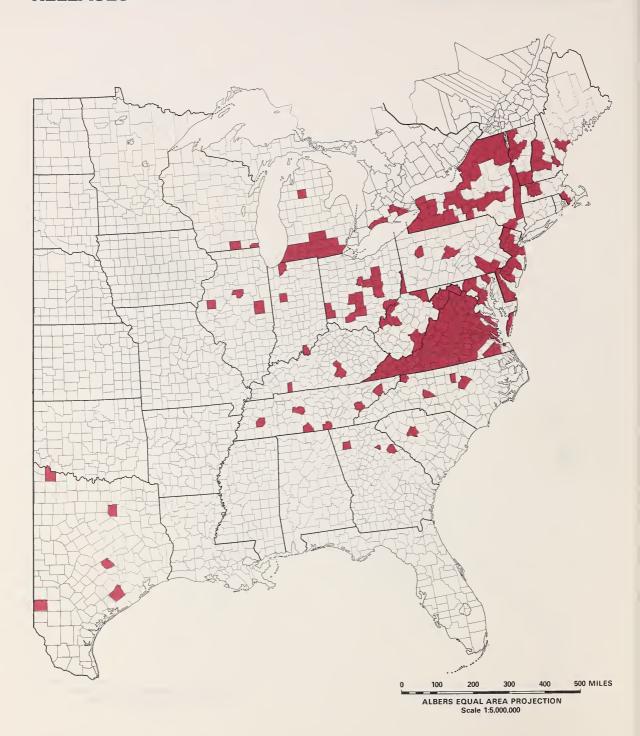
BATHYPLECTES ANURUS RELEASES (TABLE 1)



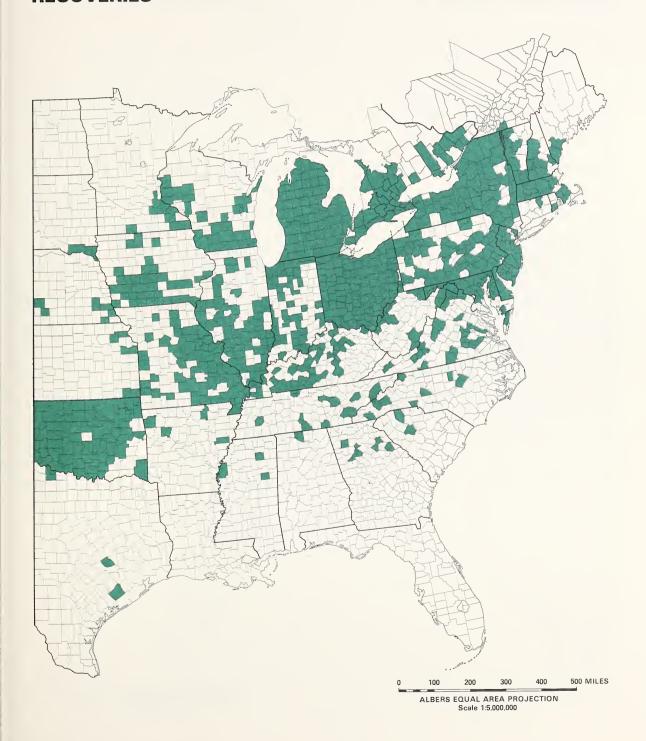
BATHYPLECTES ANURUS RECOVERIES (TABLE 1)



BATHYPLECTES CURCULIONIS RELEASES



BATHYPLECTES CURCULIONIS **RECOVERIES**



BATHYPLECTES STENOSTIGMA RELEASES (TABLE 1)



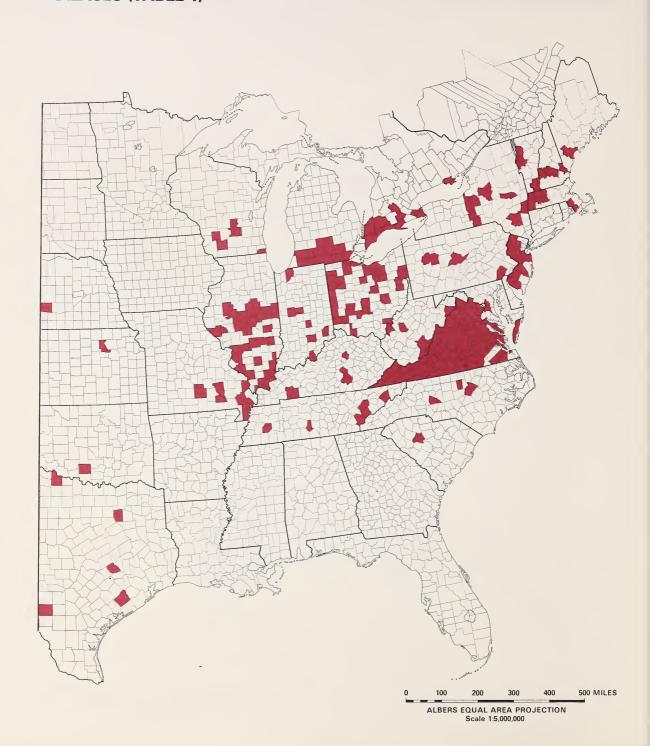
BATHYPLECTES STENOSTIGMA RECOVERIES (TABLE 1)



TETRASTICHUS INCERTUS

MAP 8

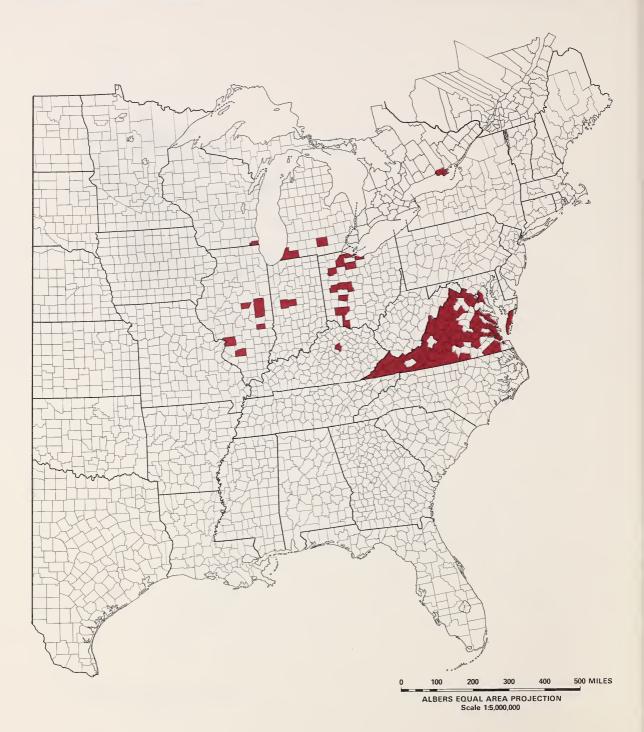
RELEASES (TABLE 1)



TETRASTICHUS INCERTUS RECOVERIES (TABLE 1)



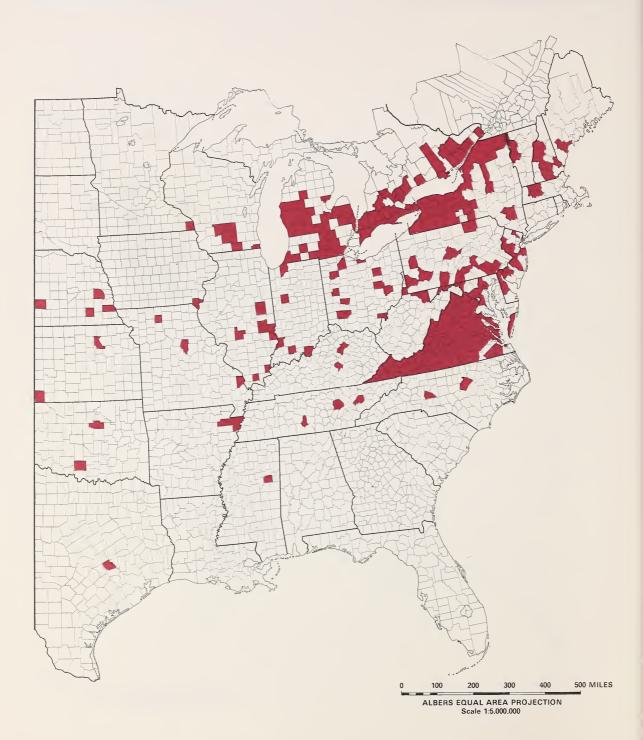
MICROCTONUS COLESI RELEASES (TABLE 1)



MICROCTONUS COLESI RECOVERIES (TABLE 1)



MICROCTONUS AETHIOPOIDES RELEASES (TABLE 1)



MICROCTONUS AETHIOPOIDES RECOVERIES (TABLE 1)



PERIDESMIA DISCUS RELEASES (TABLE 2)



TRICHOMALUS INOPS RELEASES (TABLE 2)



DIBRACHOIDES DYNASTES RELEASES (TABLE 2)



NECREMNUS LEUCARTHROS RELEASES (TABLE 2)



CAMPOGASTER EXIGUA RELEASES (TABLE 2)



MICROCTONUS STELLERI RELEASES (TABLE 2)







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